# Phylogenetic Analysis of the O-antigen Biosynthesis Genes in Vibrio cholerae

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# Introduction

#### NC\_002505.1:245142..270435 Vibrio cholerae O1 biovar El Tor str. N16961 chromosome I, complete sequence

	246 K	247 K	248 K	249 K	250 K	251 K	252 K	253 K	254 K	255 K	256 K	257 K	258 K	259 K	260 K	261 K	262 K	263 K	264 K	265 K	266 K	267 K	268 K	269 K	<u></u>
Jenes	N	V P_229899.1	C0242		VC0245 NP_229902.1		5 NP_2		/C0248 29905.1		NP_229	NP_229908.1		VC0251		VC VC0257		C0258 >			VCC	VC0260 NP_2299			
VC0241 NP_229898.2		41	NP_229		VC0244 9901.1				VC0249 P_229906.1					VC0252 NP_229909.1	,	NP_2	229913.1 0256	)913.1 56		VC0259 NP_2;	229915.2		NP_229920.	VC0264 1	C0264
rfaD NP_229897.1		397.1	VC0243 NP_229900.1				VC0247 NP_229904.1			\ NP_229907		C0250 1				NP_229912.1 VC0255					NP_229	VC0262 229918.1			
						NP	VC024 229903.1	6								NP_229911	.1					V NP_2299	/C0263 919.1		

The lipopolysaccharide (LPS) of Vibrio cholerae is a virulence factor involved in host-pathogen interactions. In particular, the O-antigen constituent of the LPS exhibits diverse genetic organization and is useful for classifying Vibrio strains and serogroups. Consequently, this provides valuable information for research into the ongoing pandemic. Our previous study developed a simple and effective bioinformatics pipeline to analyze the *wb*\* gene cluster involved in O-antigen biosynthesis. The pipeline successfully extracted these regions from publicly available, whole genome sequencing data and generated a

bootstrapped, approximately-maximum-likelihood phylogenetic tree. This follow-up study compares the *wb\** region against the genomic backbone using phylogenic methods. Results from this study facilitate the identification and analysis of horizontal gene transfer (HGT) events, particularly those involving epidemic and non-epidemic strains.

### Methods



The flowcharts illustrate the generalized pipeline for the extraction of flanked regions. The download phase issues a query to the NCBI and downloads the results as FASTA files. The query phase makes a BLAST database to search for hits to the two flank sequences. The extraction phase relies on two in-house scripts that produce an entry\_batch file and download a corresponding GenBank file representative of the extracted regions. The entry\_batch lists the header, range, and strand of each region. The GenBank is useful for extracting genes and metadata, particularly serogroup information. The alignment phase removes duplicate sequences, producing a non-redundant FASTA file and tab-delimited file matching the representative sequence to duplicates. Next, the MAFFT tool generates a multiple alignment file. The FastTree program performs approximate, maximum-likelihood estimation with bootstrapping. The sed command re-labels the taxon identifiers and MEGA renders the tree.



# Conclusion

The pipeline successfully extracted the *wb*\* regions. The corresponding tree consists of a large clade of O1 taxa and a small clade of O139 taxa. The rest of the tree consists of other serogroup taxa. Two O1 serogroup taxa occur within this portion.

Multiple Non-O1/O139 taxa were observed in the tree. Horizontal gene transfer may be responsible for the occurrence of conserved flanking genes, *gmhD* and *rjg*, across the serogroups. Additional analysis is required to investigate the genes that constitute the region.

Metadata is critical for conducting additional analysis. A semi-automated process extracted serogroup data. Considerable manual effort was required to obtain as much information as possible. This is due to user error and lack of standards when submitting sequence feature qualifiers to GenBank. Thus, further literature review is required to obtain missing serogroup information.

Further investigation aims to compare the evolution of the genomic backbone with the *wb\** region. This includes phylogenetic analysis of the *wb\** regions for each serogroup separately. The generalized conserved region pipeline may be applied to the study of other gene clusters.



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# Acknowledgements

Approximately-maximum-likelihood phylogenetic tree of the *wb\** region. Each label groups the regions Special thanks to the Big Data Laboratory at Noblis, Inc., especially Mitch Holland for reviewing this of serogroups with 100% identity: label = N<sub>k</sub>S<sub>k</sub>, N = number of taxa, S = serogroup, k = serogroup index. poster. Thank you, SFAF 2016!